

Single-cell Analysis and its Application in Virology

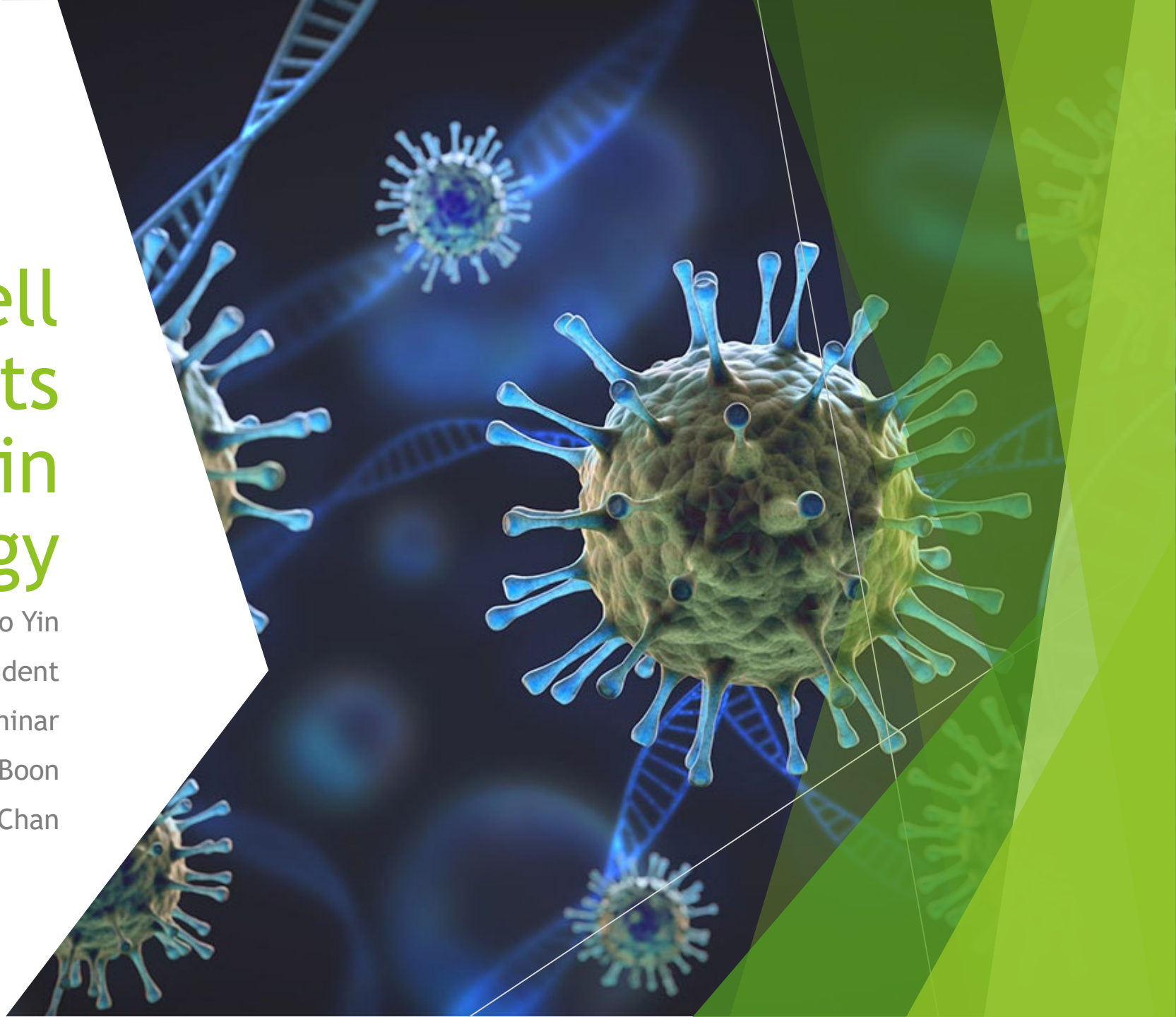
Peter Luk Ho Yin

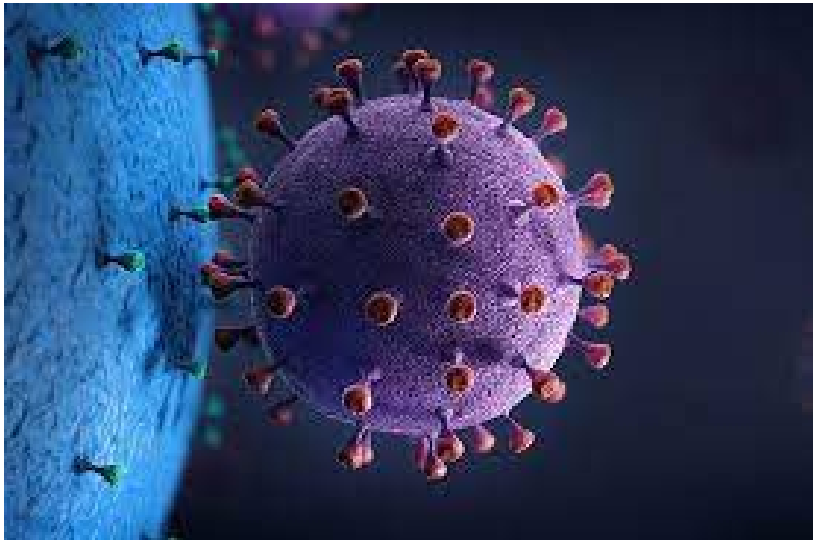
2nd year PhD student

Joint Graduate Seminar

Supervisor: Dr Siaw Shi Boon

Co-supervisor: Prof. Paul Chan



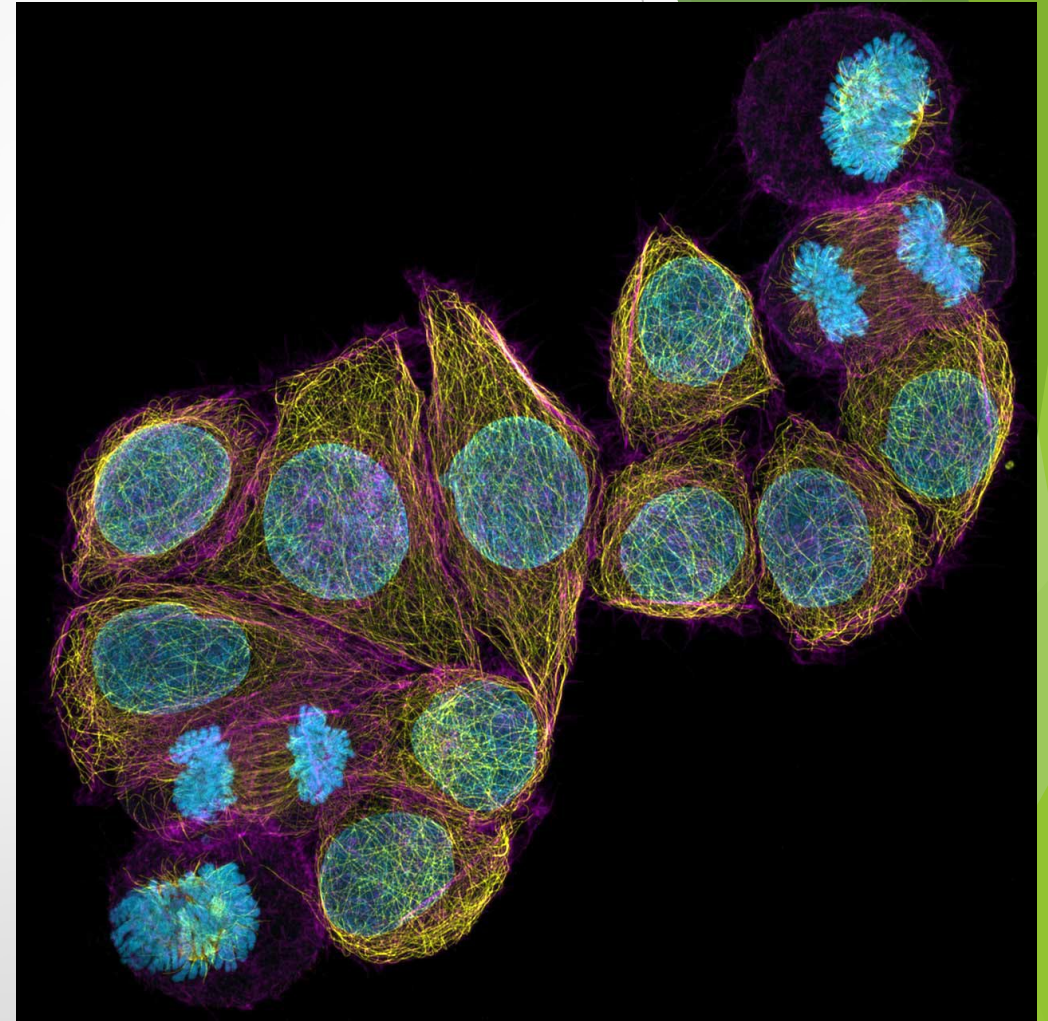


Outline

- ▶ Cell heterogeneity
- ▶ Isolation of single cells
- ▶ Amplification of genetic material
- ▶ Application of single cell in virology
- ▶ Conclusion

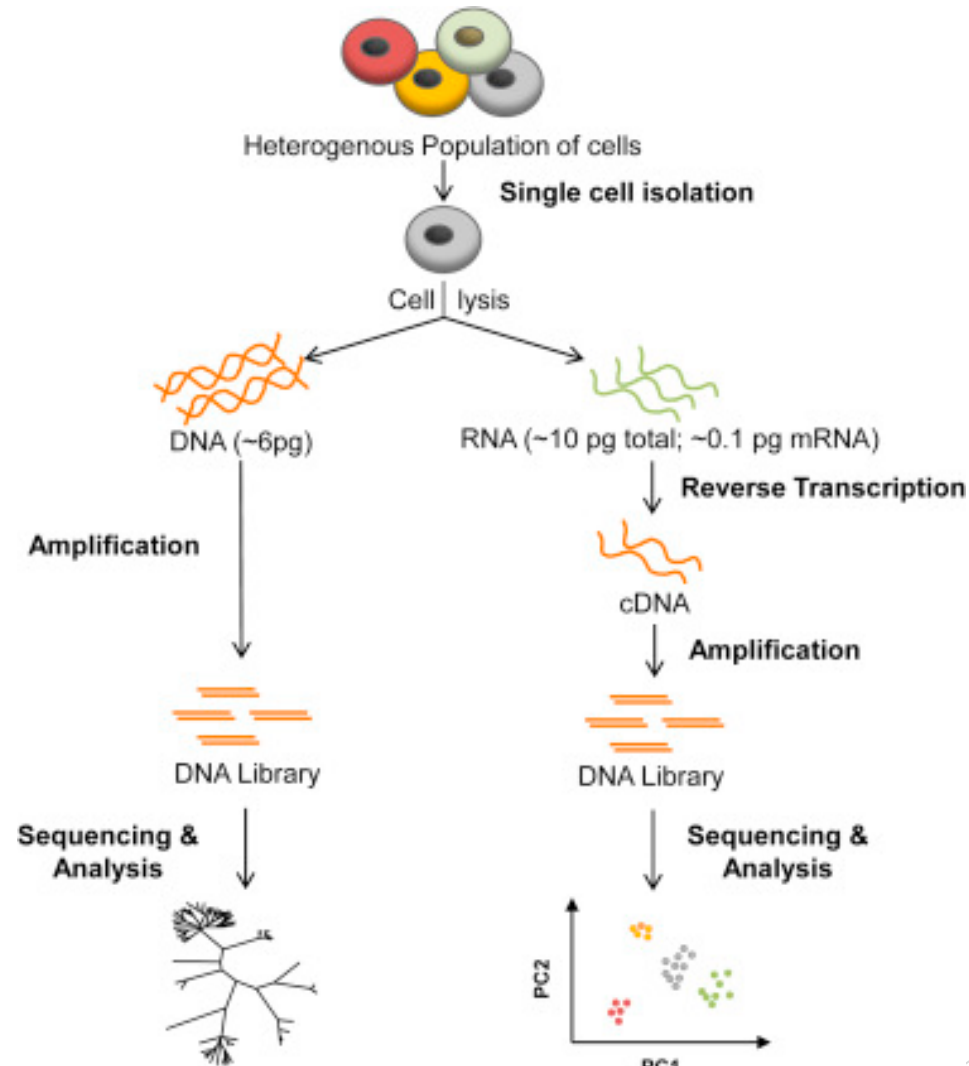
Cellular Heterogeneity

- ▶ Most of our current biological knowledge based on ensemble measurements
- ▶ Cell-to-cell in a population differs:
 - Identity (Cell types, subpopulation/ lineage)
 - state/process (Cell cycle, circadian rhythm)
 - stochastic variation



Single-cell (Sc) analysis

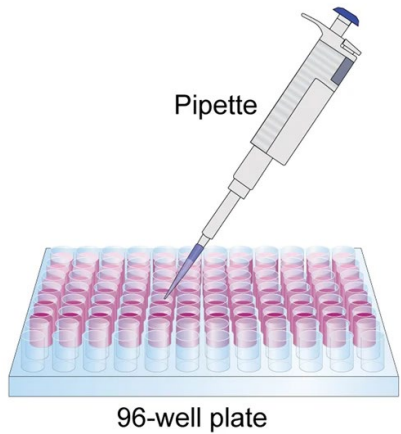
- ▶ Powerful tool to study cell heterogeneity through analysis of whole genome and transcriptome of individual cell
- ▶ Identify minority sub-population
- ▶ Discover unique characteristics of individual cells
- ▶ Proven useful in cancer, immunology, embryology and microbiology



S Rato et al., Virus Research(2017)

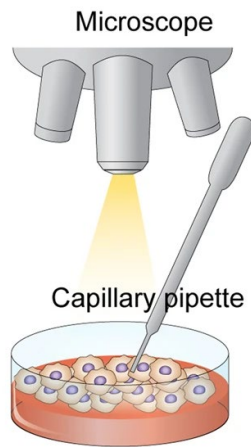
Single-cell isolation techniques

a



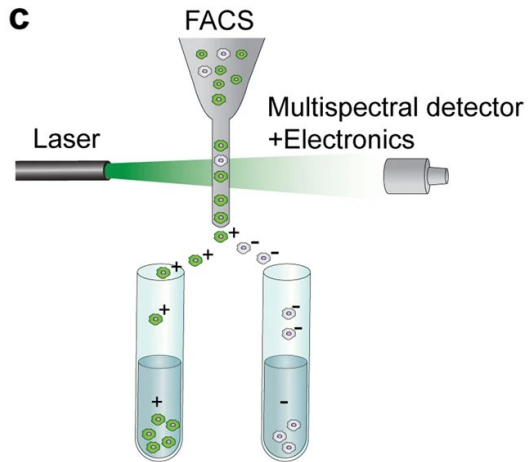
Limiting dilution

b



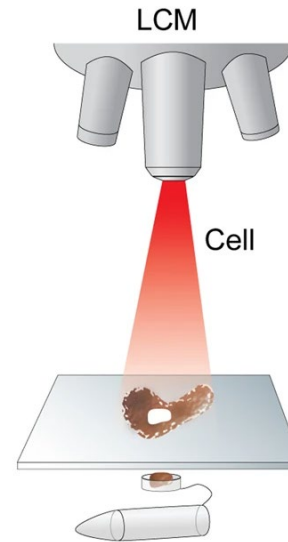
Micromanipulation

c



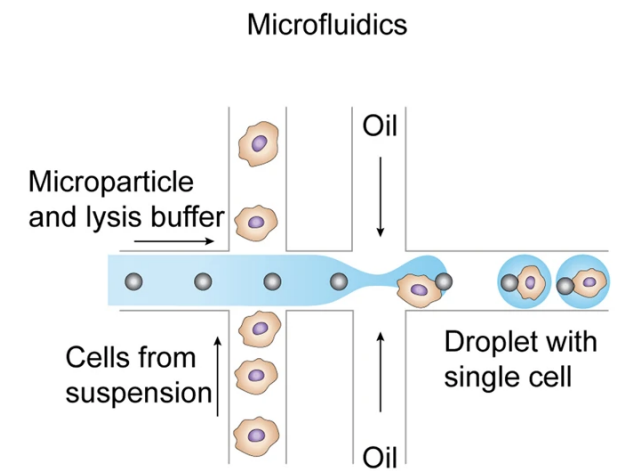
Fluorescence-activated cell sorting (FACS)

d

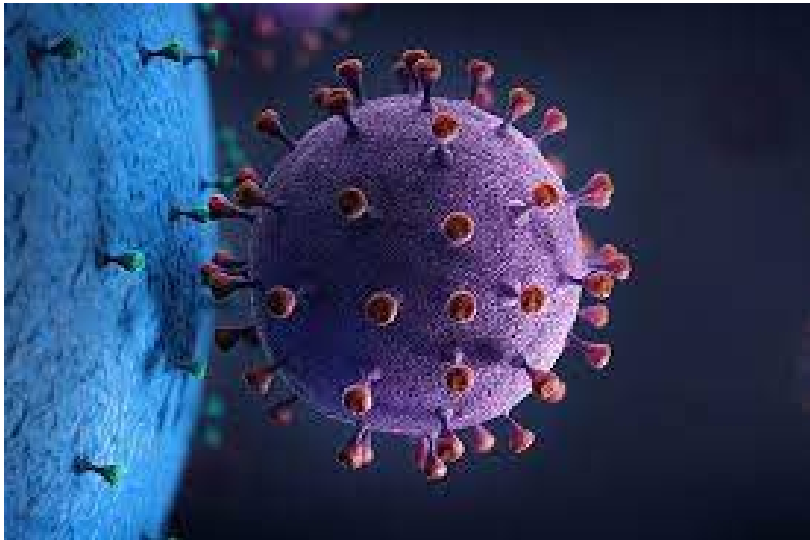


Laser capture microdissection (LCM)

e



Microdroplet-based microfluidics



Outline

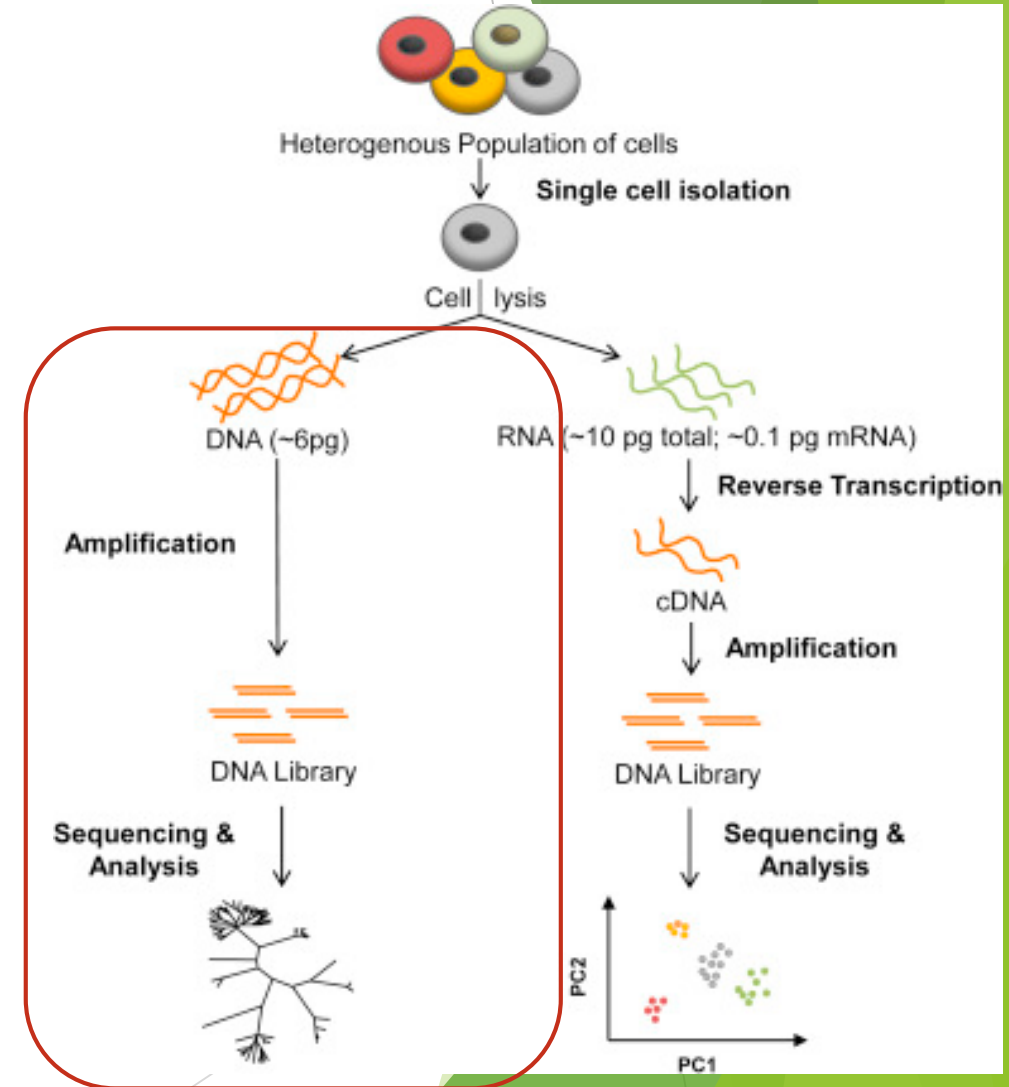
- ▶ Cell heterogeneity
- ▶ Isolation of single cells
- ▶ **Amplification of genetic material**
- ▶ Application of single cell in virology
- ▶ Conclusion

Whole genome amplification (WGA)

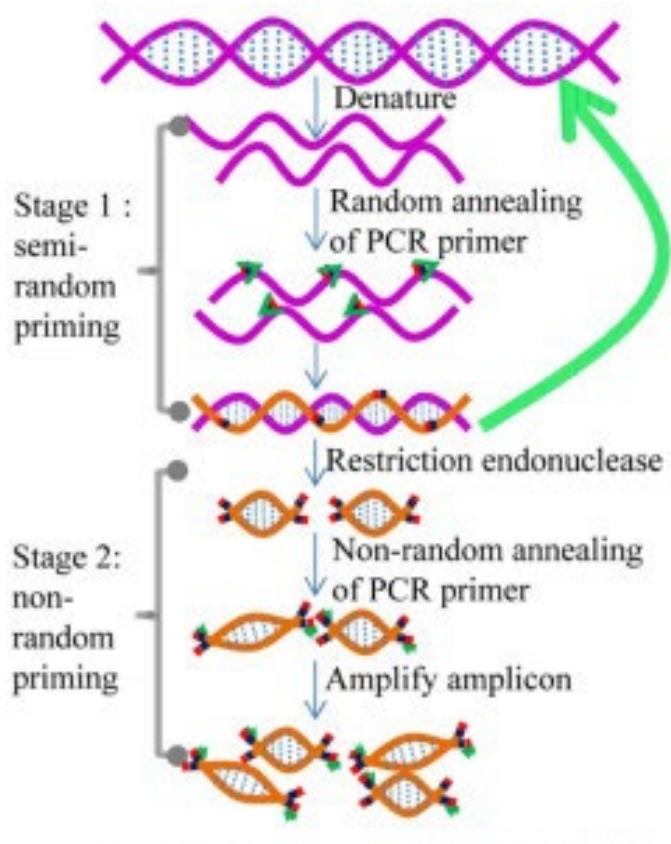
- ▶ Normal diploid human cell: 6-7 pg DNA, inadequate for genomic sequencing

Amplification step:

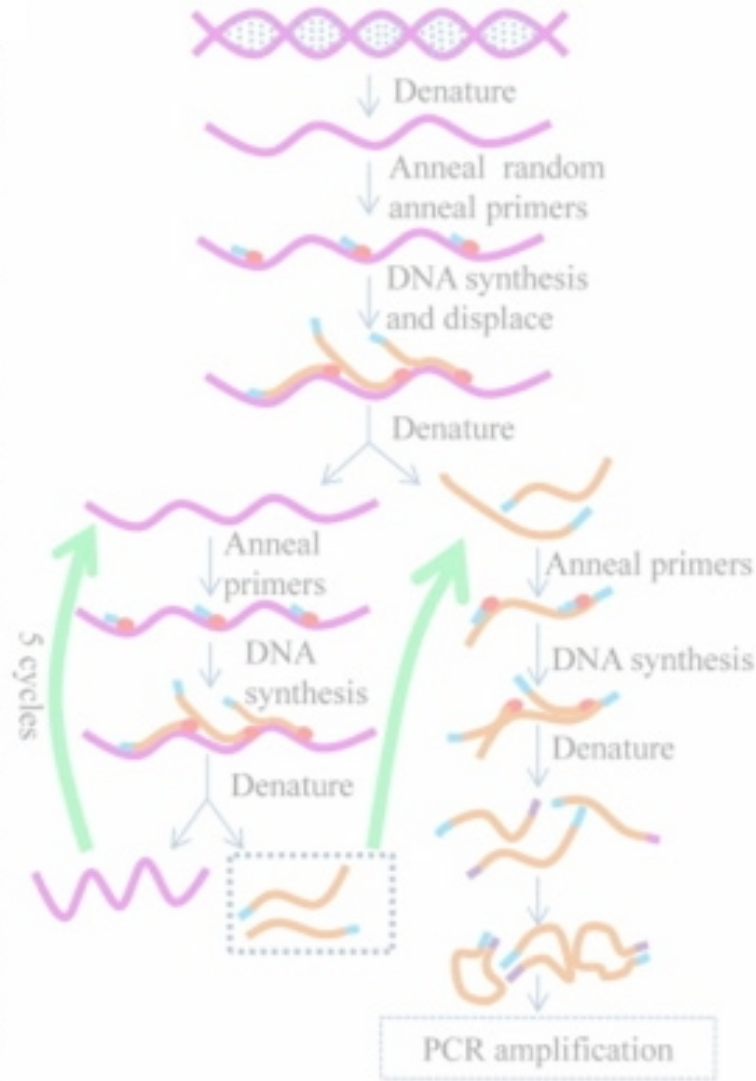
- ▶ degenerate-oligonucleotide-primed PCR (DOP-PCR)
- ▶ multiple annealing and looping-based amplification cycles (MALBAC)
- ▶ multiple-displacement-amplification (MDA)



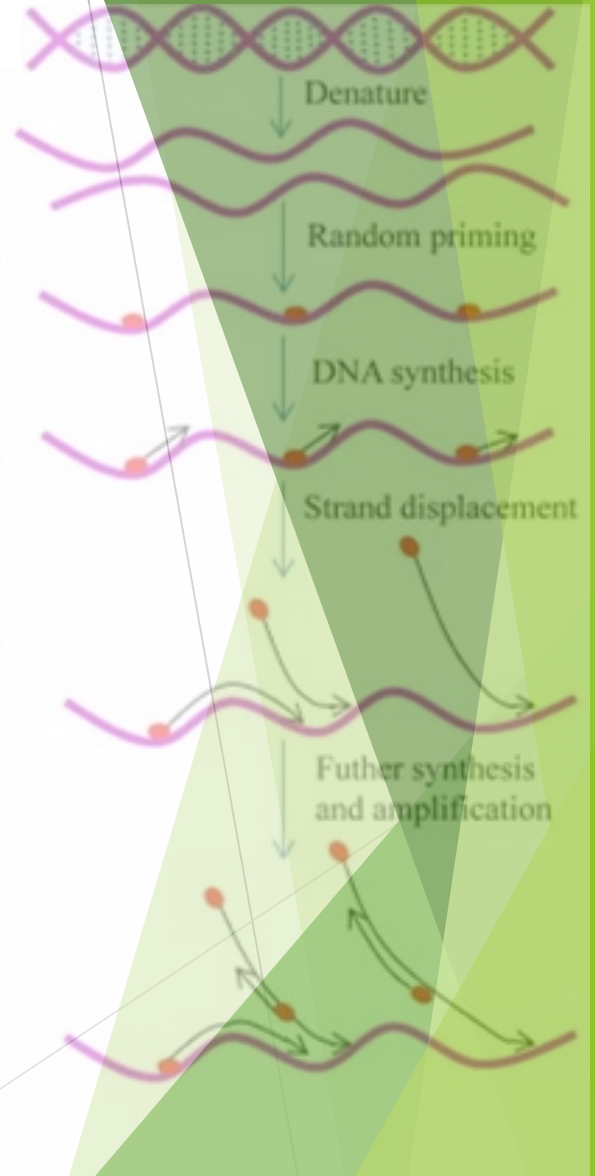
Degenerate oligonucleotide primed (DOP)-PCR



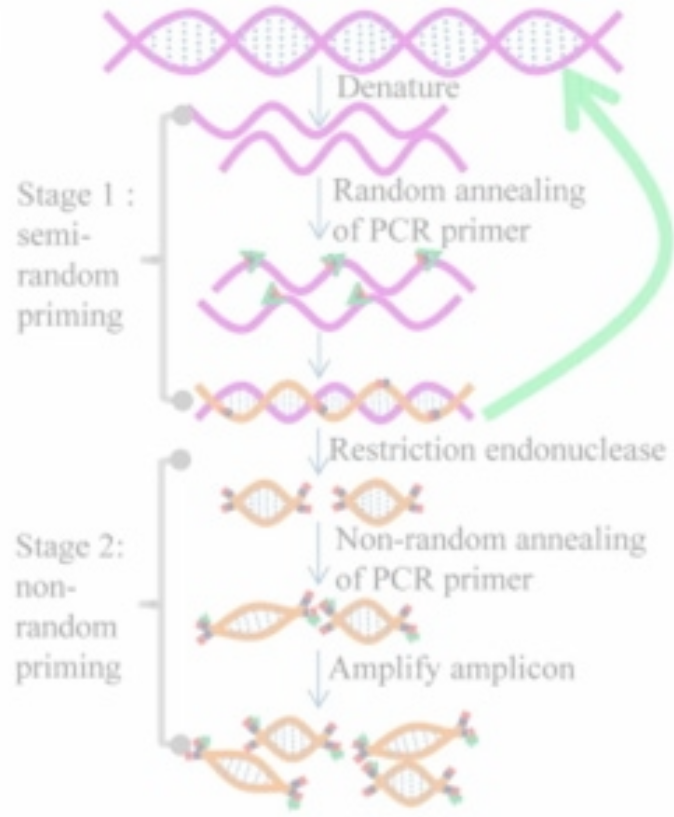
Multiple Annealing and Looping-Based Amplification Cycles (MALBAC)



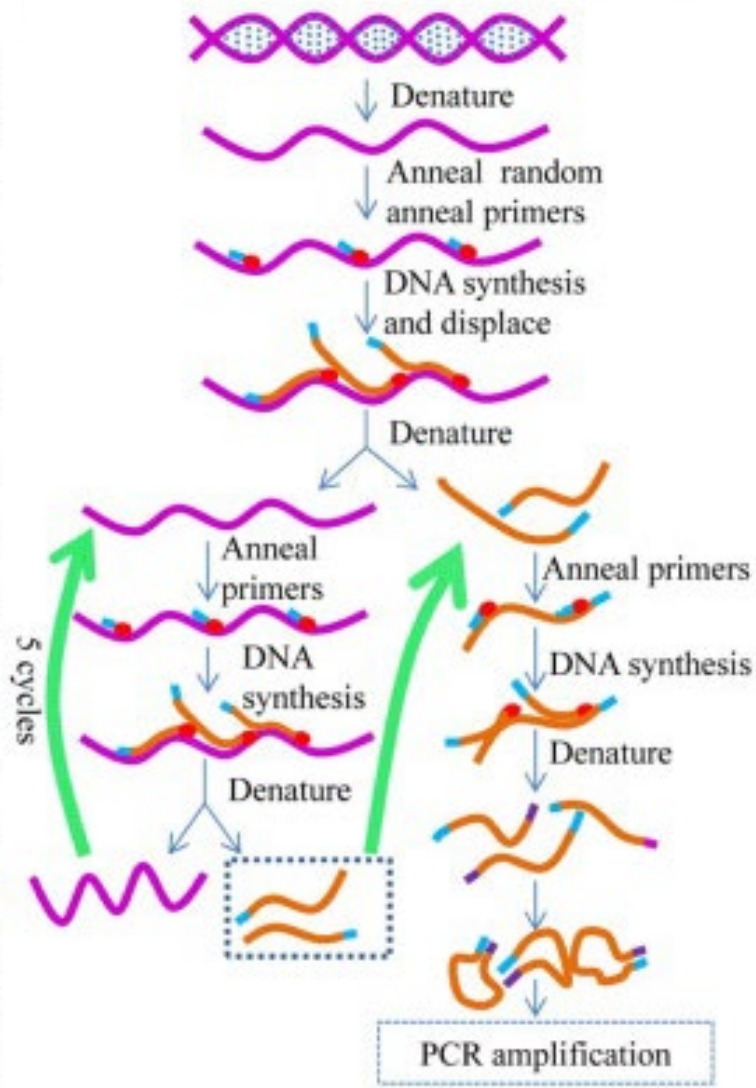
Multiple displacement amplification (MDA)



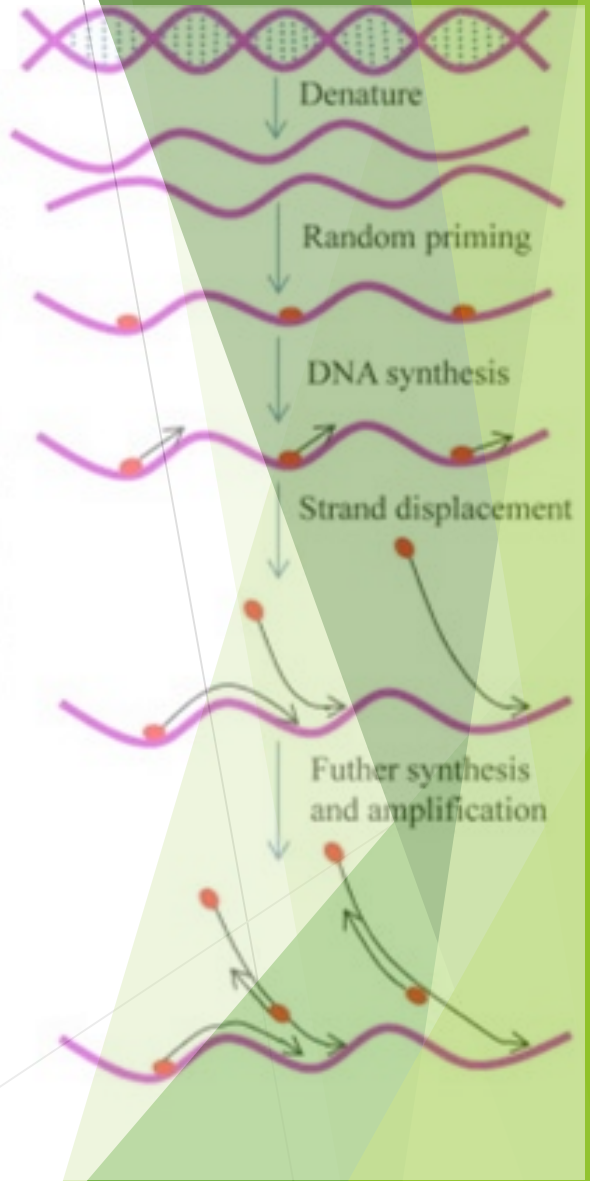
Degenerate oligonucleotide primed (DOP)-PCR



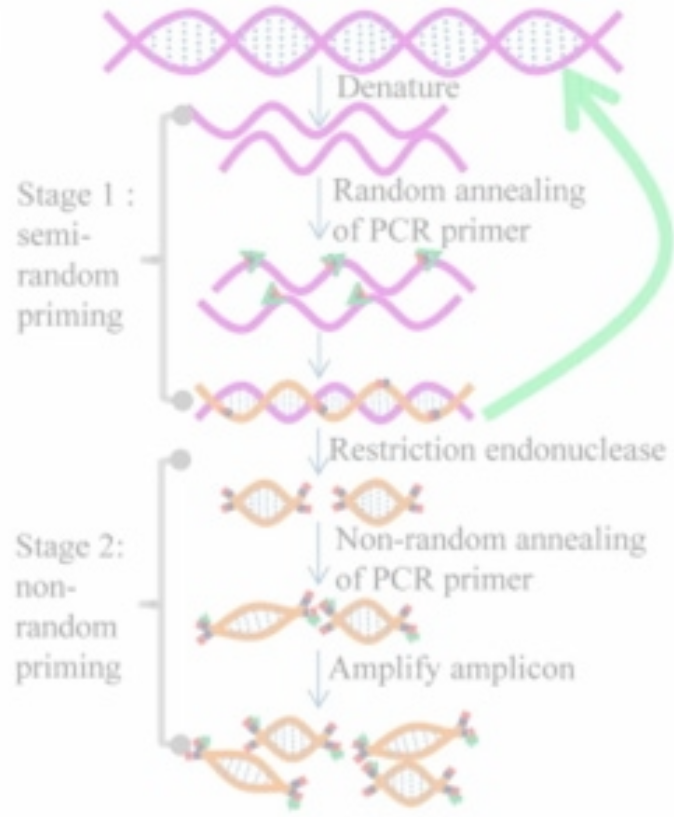
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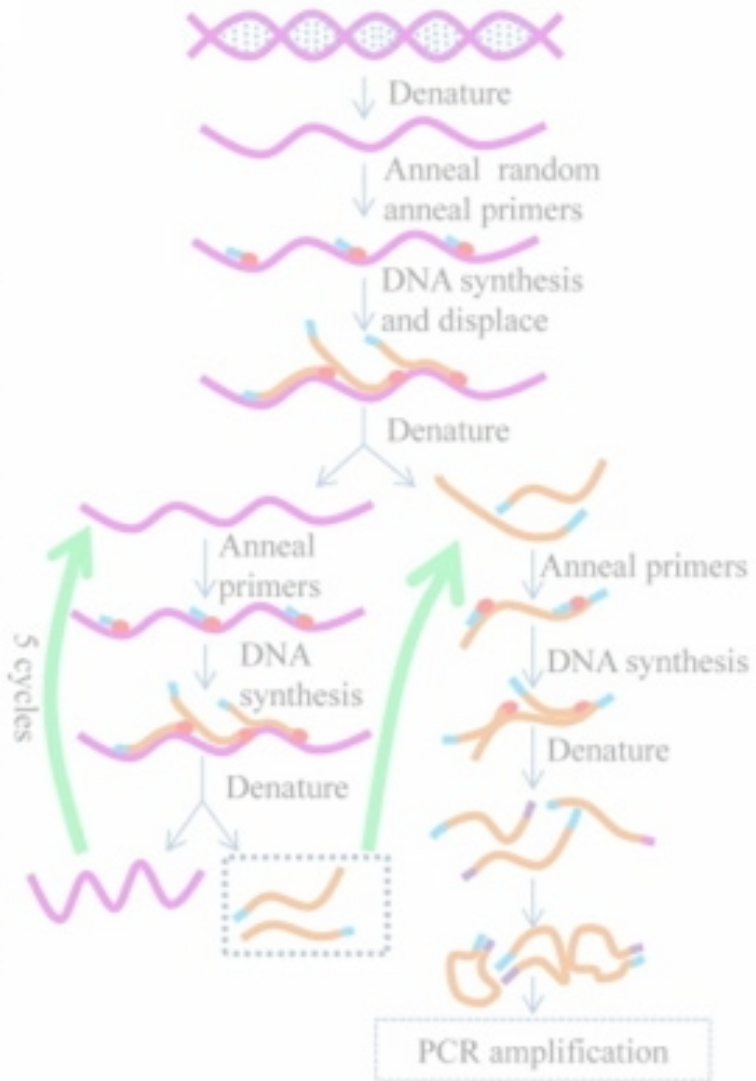
Multiple displacement amplification (MDA)



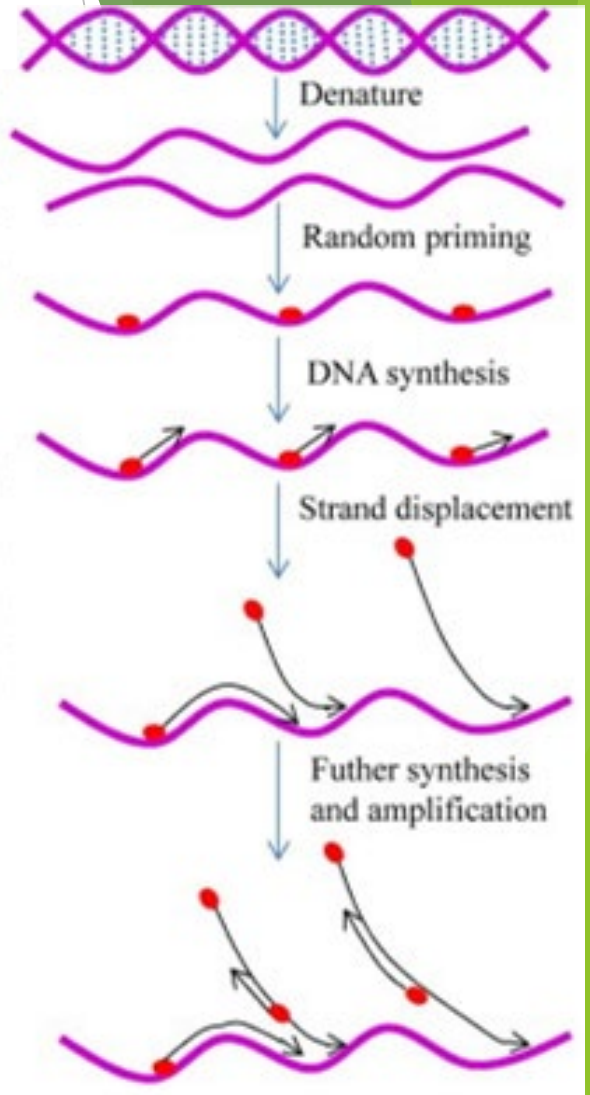
Degenerate oligonucleotide primed (DOP)-PCR



Multiple Annealing and Looping-Based Amplification Cycles (MALBAC)

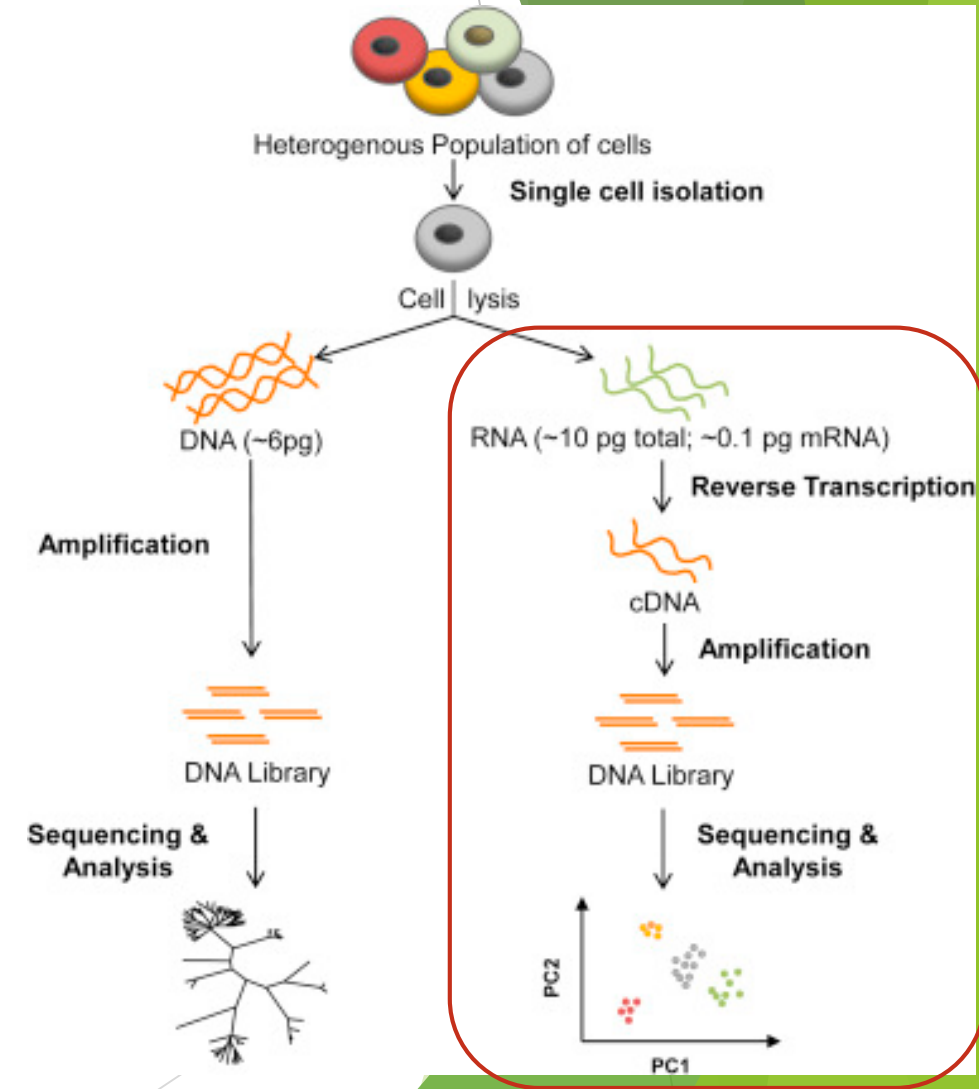


Multiple displacement amplification (MDA)



Whole transcriptome amplification (WTA)

- ▶ Amount of RNA in single cell inadequate
- ▶ WTA generate cDNA library for single cell transcriptome sequencing
- ▶ Traditional/ modified PCR
- ▶ T7-*in vitro* transcription (IVT)
- ▶ Phi29 DNA polymerase-mediated RNA amplification (PMA)

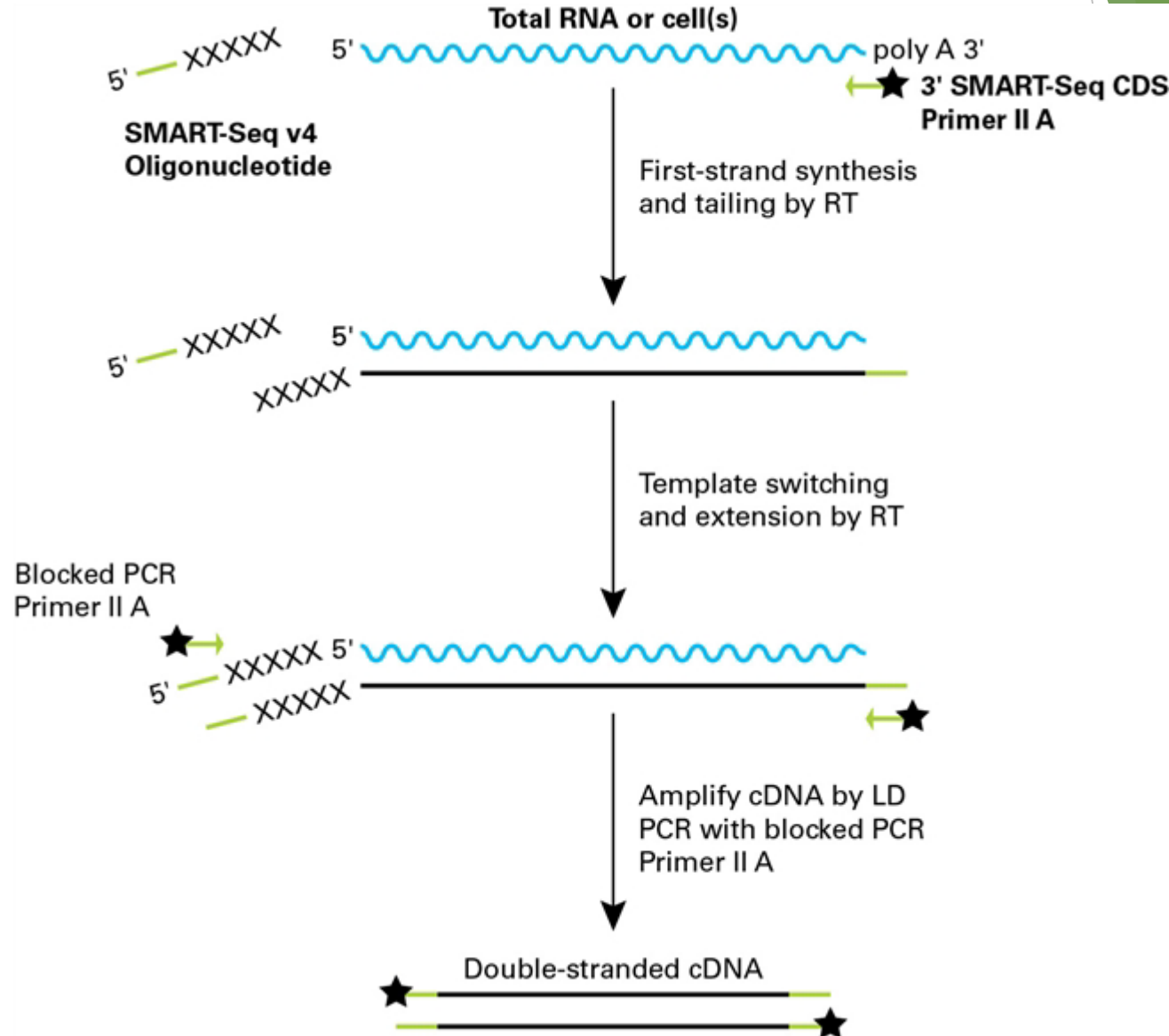


S Rato et al., Virus Research(2017)

Modified PCR

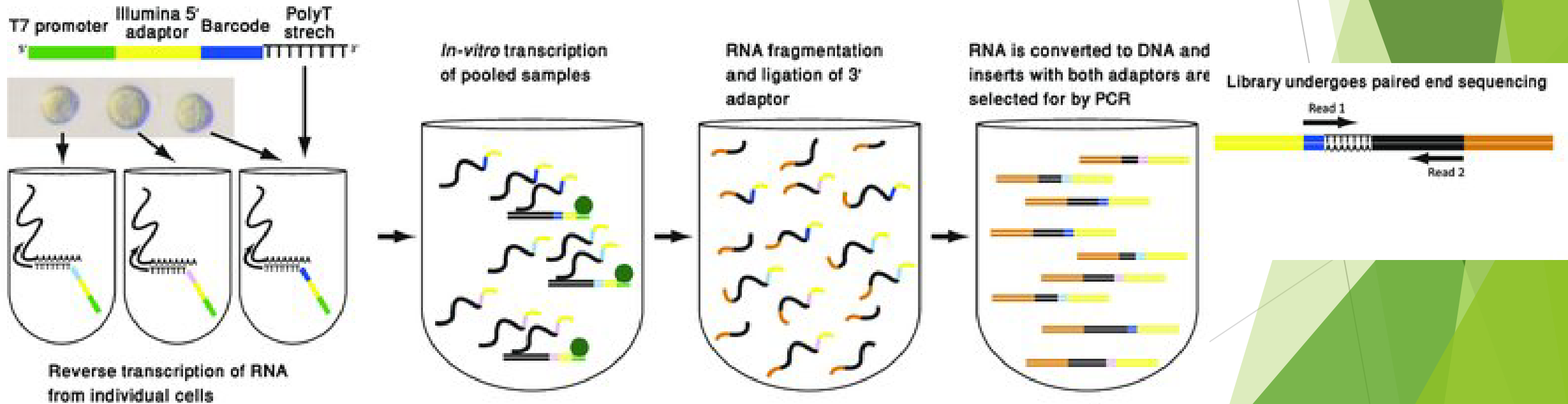
SMART techniques

“Switching Mechanism At the 5' end of the RNA Transcript”

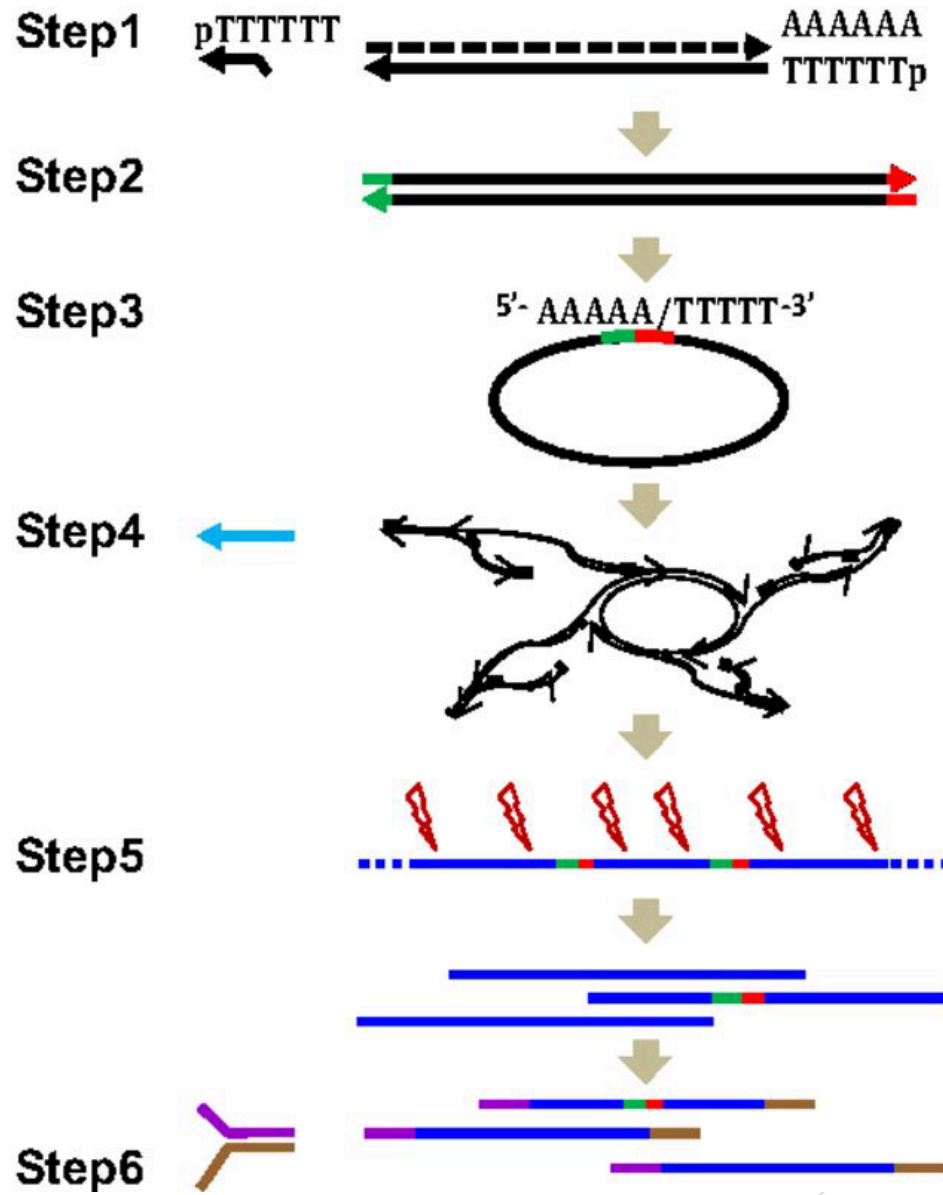


T7-*in vitro* transcription (IVT)

- ▶ Different than PCR-based method (different design of reverse transcription)
- ▶ CEL-seq (cell expression by linear amplification and sequencing)

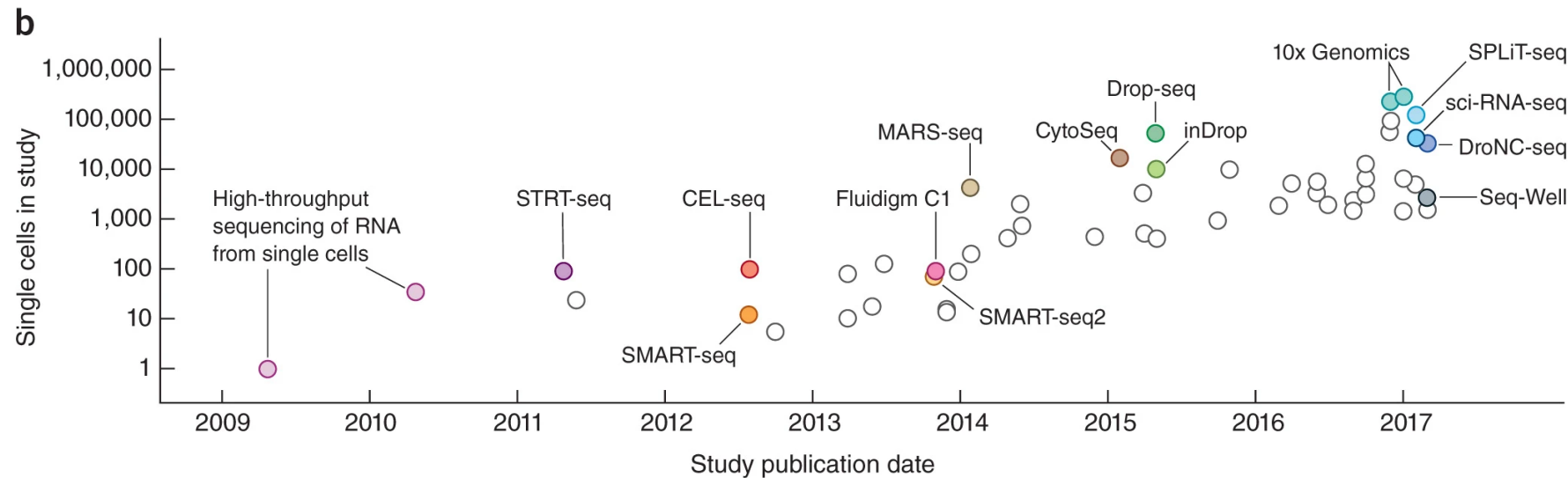
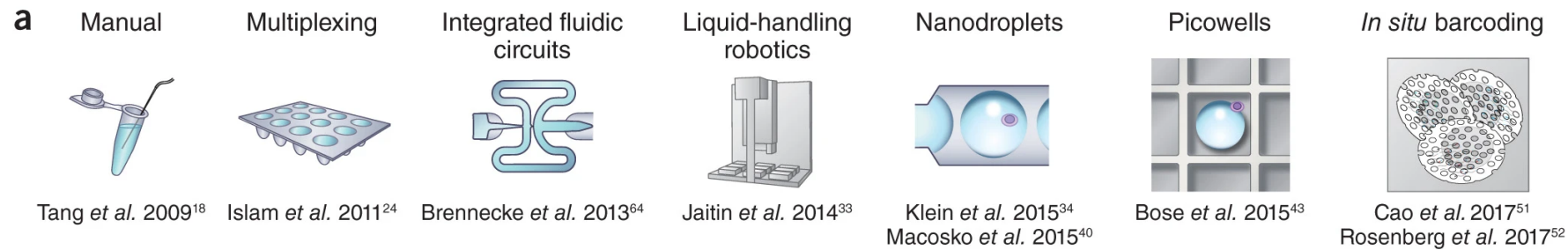


Phi29-mRNA amplification (PMA)



scRNA-seq platforms

- ▶ Tremendous growth of commercial scRNA-seq platforms
- ▶ ↑ cell number profiled, accuracy, sensitivity
- ▶ ↓ reagents, cost



Single-cell technology in Virology

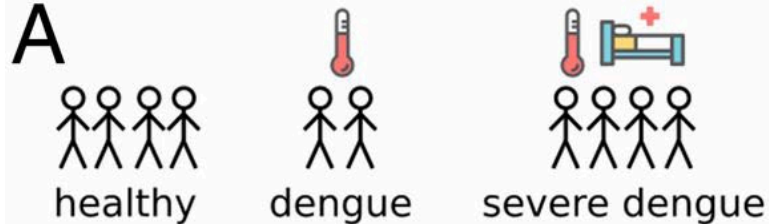
- ▶ Virus dependent on host cell to replicate
- ▶ Heterogeneity of host cell reflects in viral infection outcome
- ▶ 100% infected cells is difficult
- ▶ (cellular heterogeneity/ virus particles)
- ▶ SCs allows joint analysis of virus replication and host cell environment
- ▶ Virus/Cell-based: Viral replication, Virus-induced cellular response/ transcriptome, infection outcome

Virus-inclusive single-cell RNA sequencing reveals the molecular signature of progression to severe dengue

Fabio Zanini^{a,1}, Makeda L. Robinson^{b,c,1}, Derek Croote^a, Malaya Kumar Sahoo^d, Ana Maria Sanz^e, Eliana Ortiz-Lasso^f, Ludwig Luis Albornoz^f, Fernando Rosso^{e,g}, Jose G. Montoya^c, Leslie Goo^h, Benjamin A. Pinsky^{c,d}, Stephen R. Quake^{a,h,i,2}, and Shirit Einav^{b,c,2}

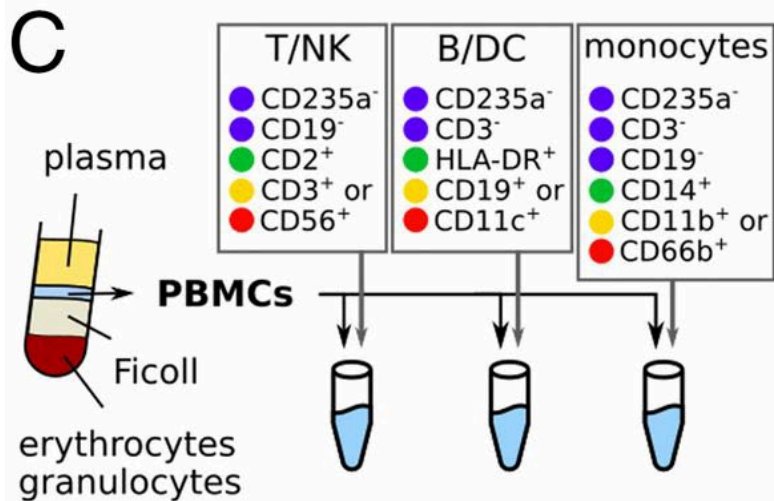
^aDepartment of Bioengineering, Stanford University, Stanford, CA 94305; ^bDepartment of Microbiology and Immunology, Stanford University School of Medicine, Stanford, CA 94305; ^cDivision of Infectious Diseases and Geographic Medicine, Department of Medicine, Stanford University School of Medicine, Stanford, CA 94305; ^dDepartment of Pathology, Stanford University School of Medicine, Stanford, CA 94304; ^eClinical Research Center, Fundación Valle del Lili, Cali 760026, Colombia; ^fPathology and Laboratory Department, Fundación Valle del Lili, Cali 760026, Colombia; ^gDivision of Infectious Diseases, Department of Internal Medicine, Fundación Valle del Lili, Cali 760026, Colombia; ^hChan Zuckerberg Biohub, San Francisco, CA 94158; and ⁱDepartment of Applied Physics, Stanford University, Stanford, CA 94305

Contributed by Stephen R. Quake, October 24, 2018 (sent for review August 10, 2018; reviewed by Katja Fink and Alex K. Shalek)

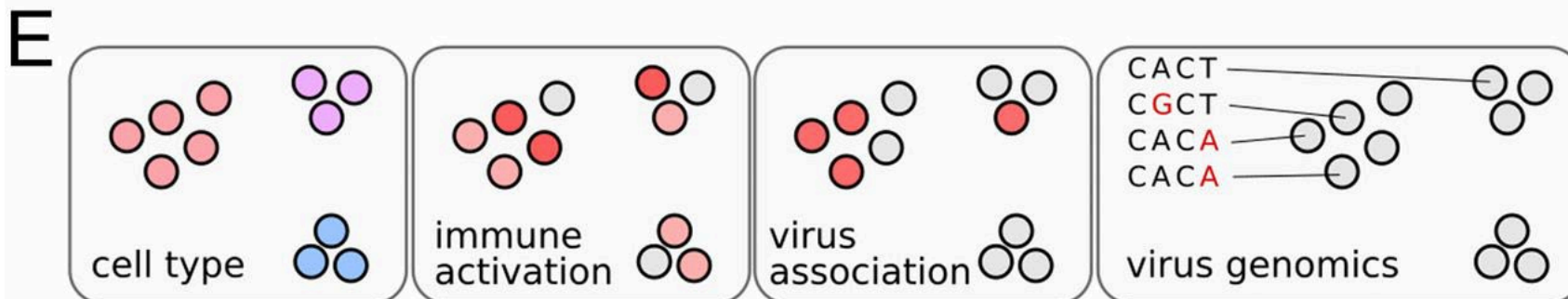
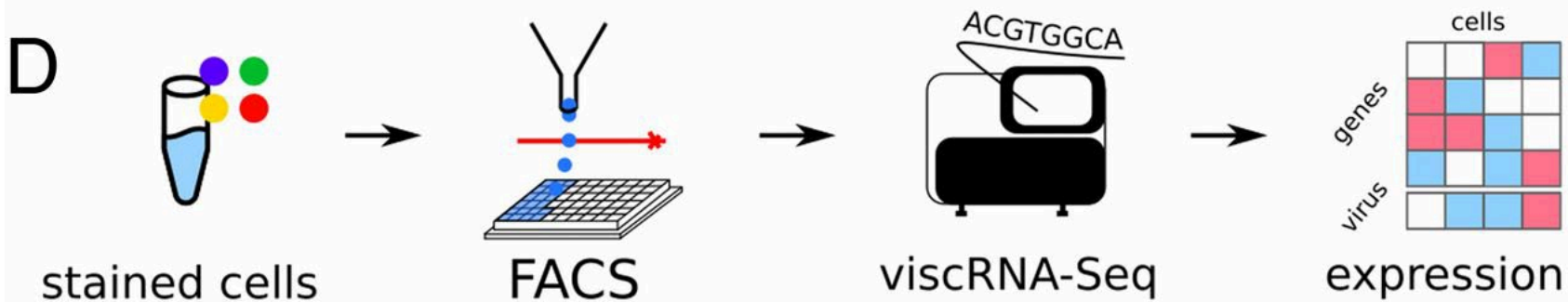


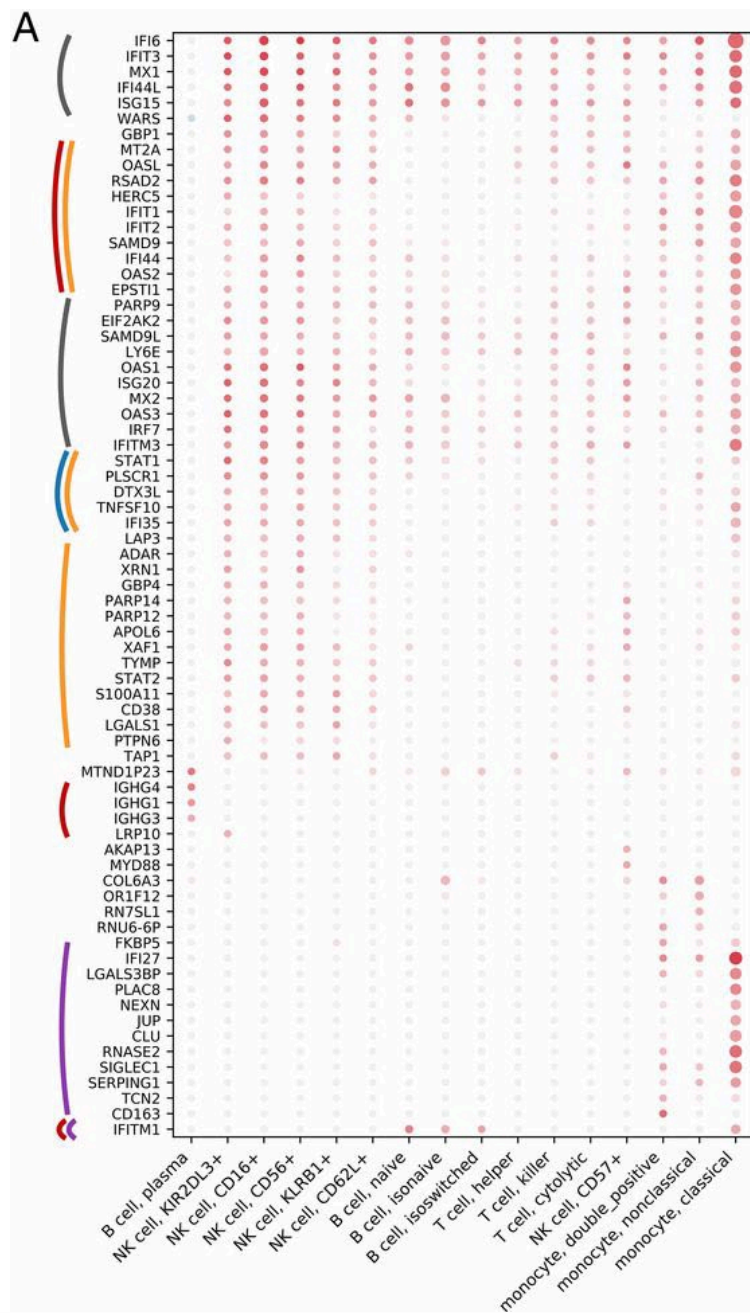
B

subject	diagnosis	serotype	viral load*
3-013-1	healthy	N.A.	0
3-027-1	healthy	N.A.	0
3-018-1	healthy	N.A.	0
3-006-1	healthy	N.A.	0
1-008-1	dengue	4	8×10^3
1-020-1	dengue	1	1×10^6
1-013-1	severe	4	9×10^3
1-026-1	severe	3	9×10^8
1-010-1	severe	3	6×10^5
1-036-1	severe	3	5×10^7



* copies / ml of plasma

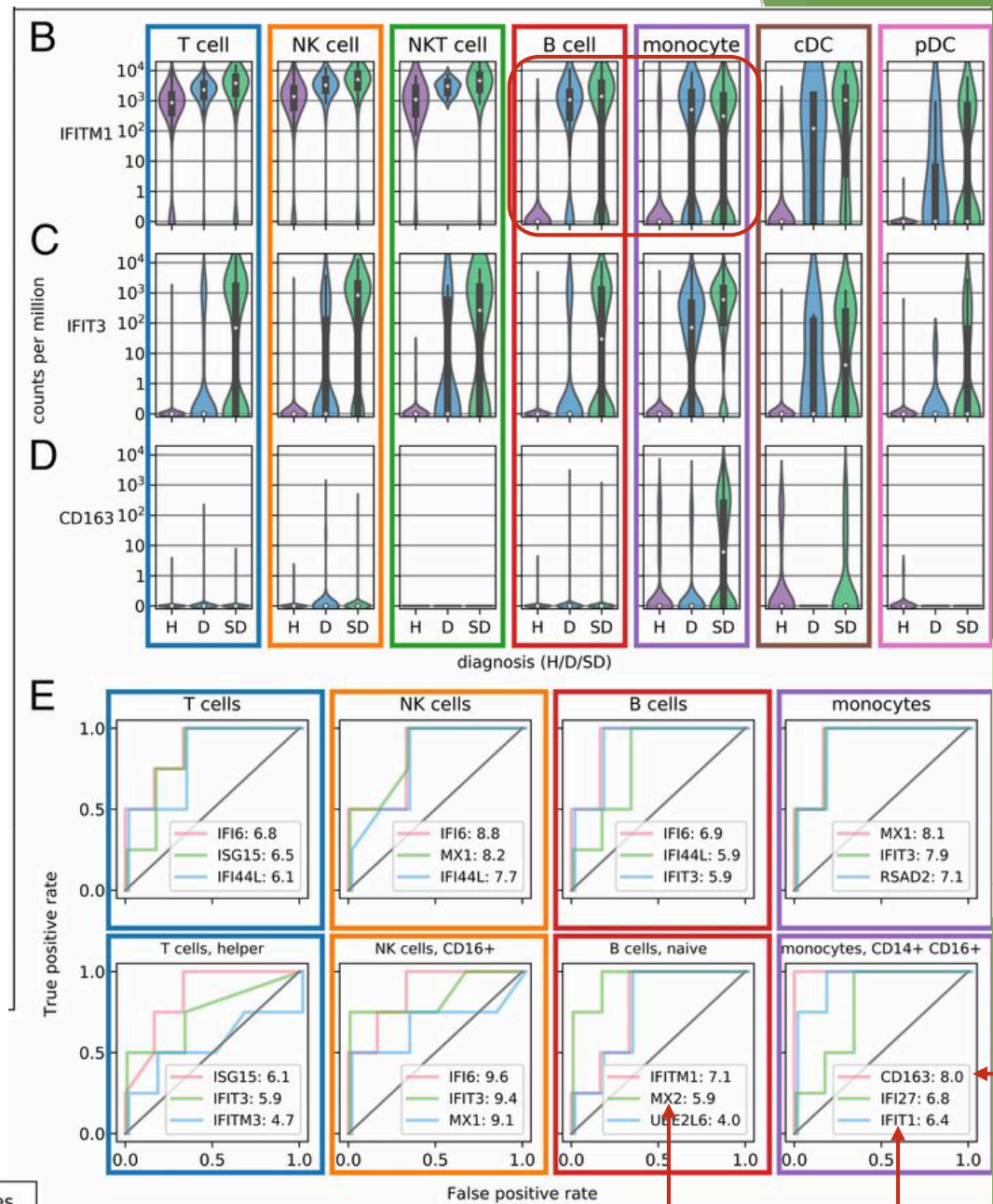




Overexpression specific to: (red) B cells (blue) T cells (orange) NK cells (purple) monocytes (grey) all cell types

Average fold change in severe dengue: ○ no change ○ ~2 to 5 fold ● ~10 fold or more

Kolmogorov Smirnov test between single cell distributions: ● less significant ● more significant










ARTICLE

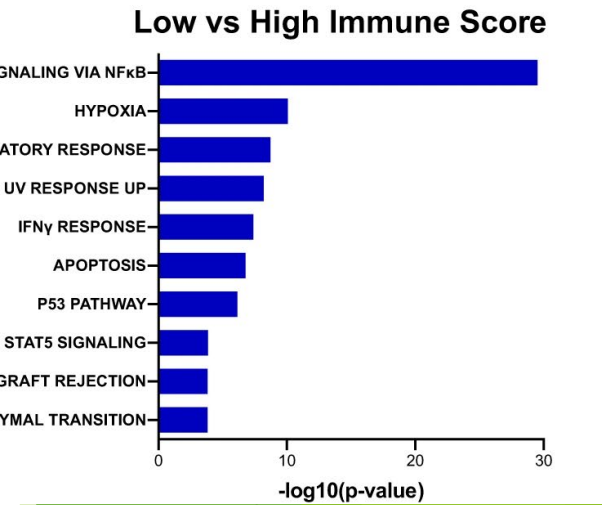
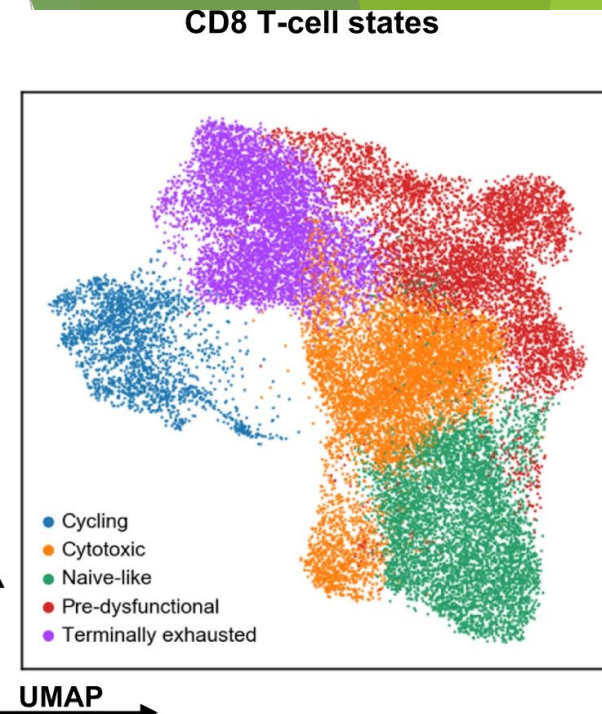
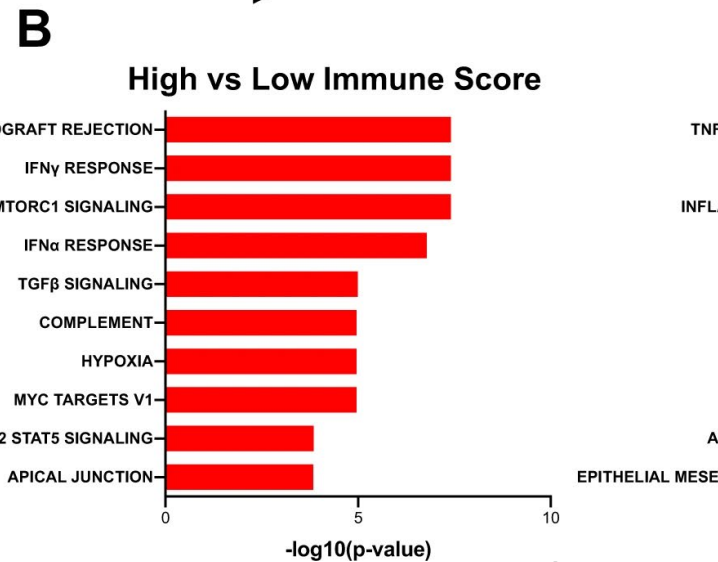
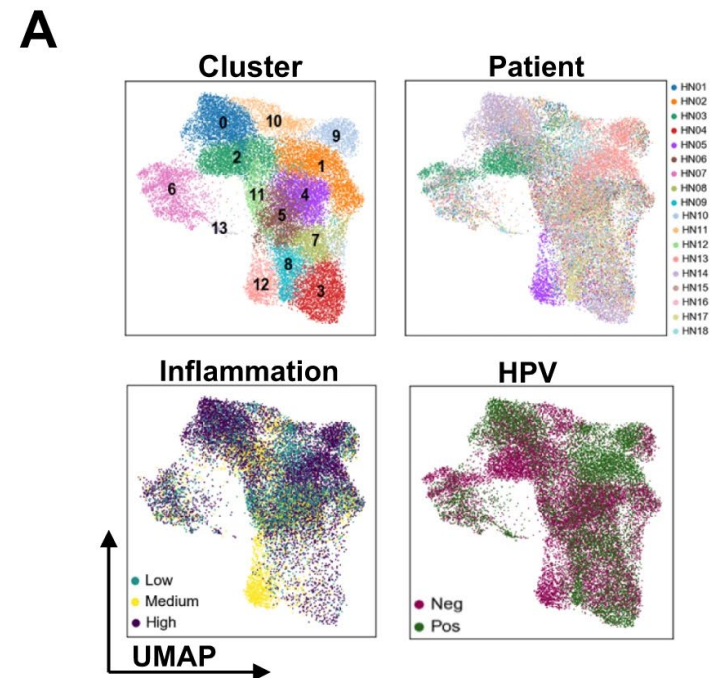
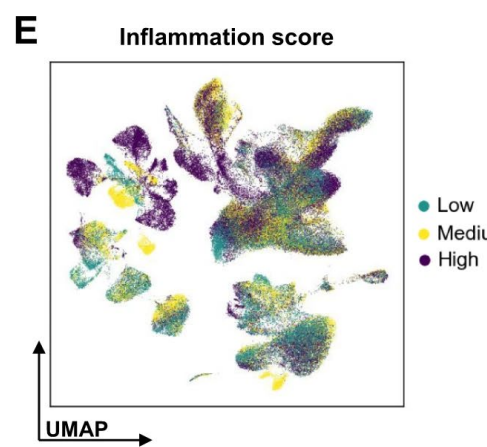
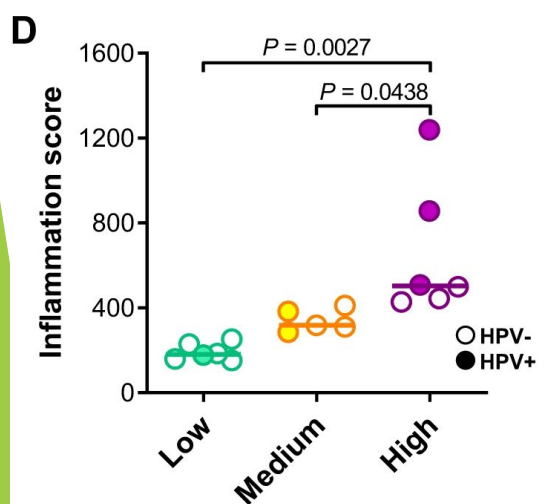
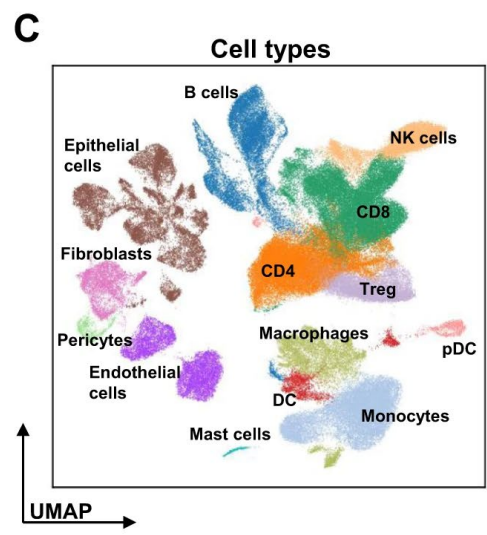
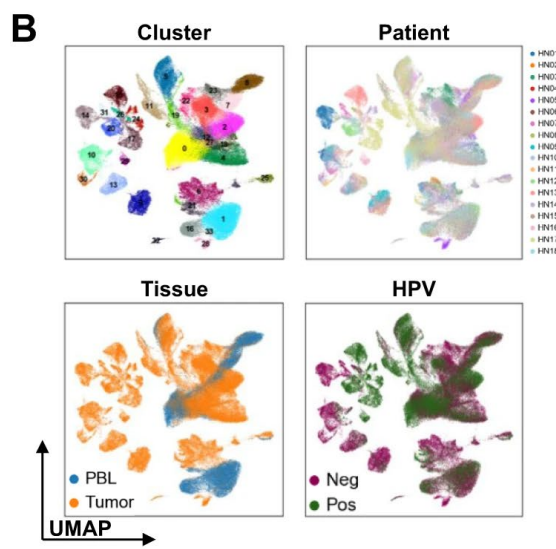
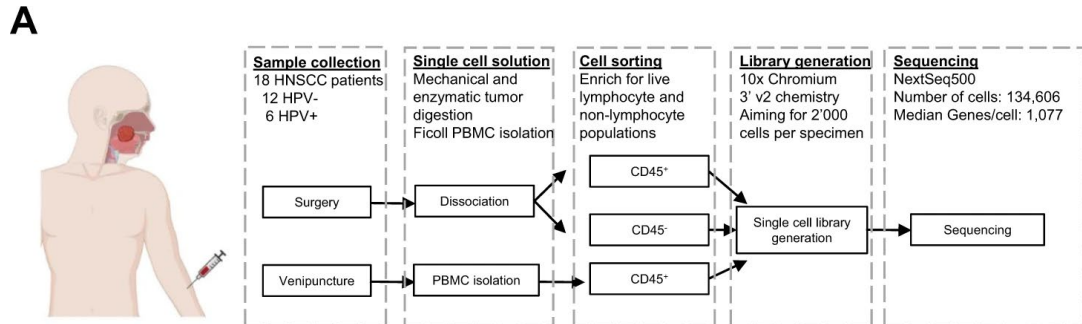


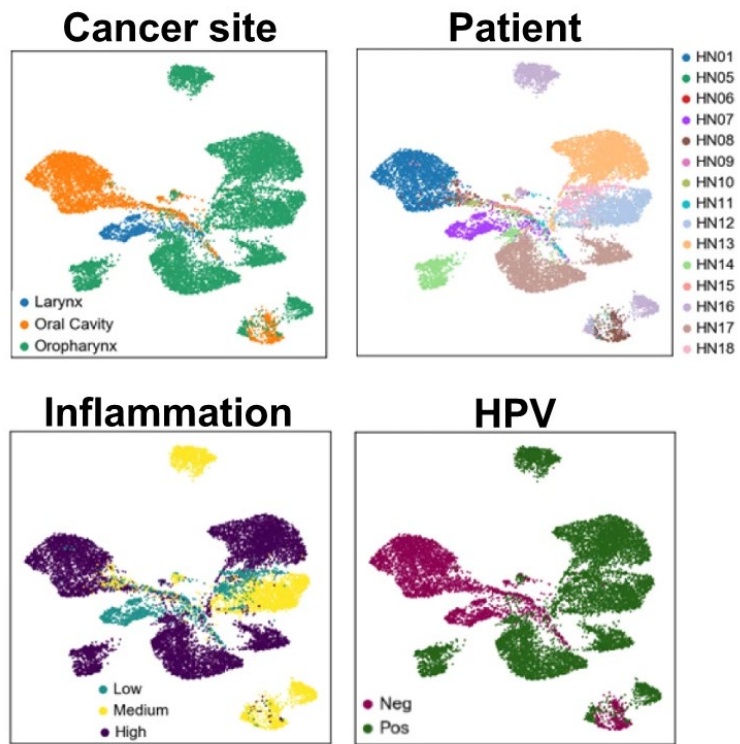
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OPEN

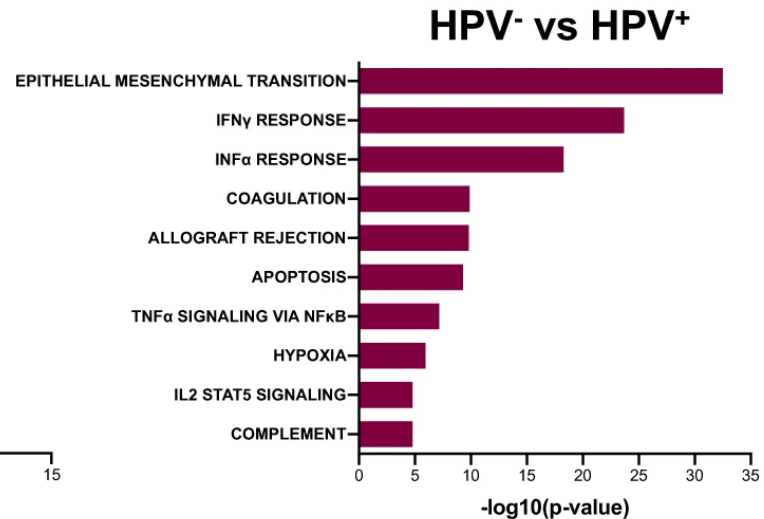
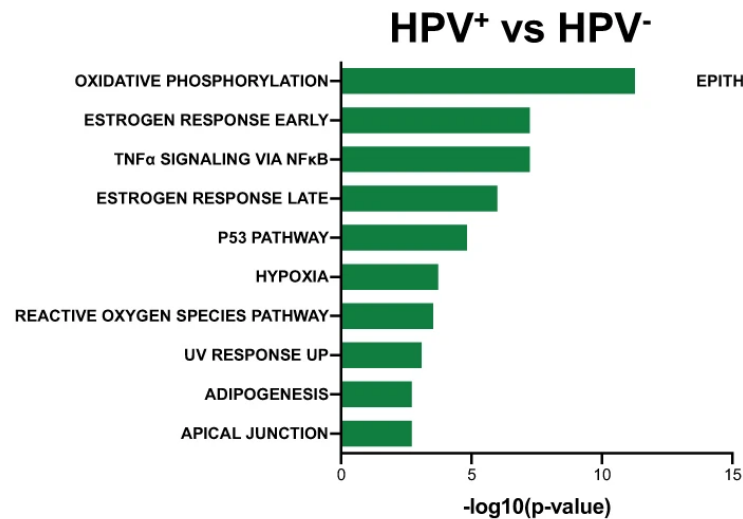
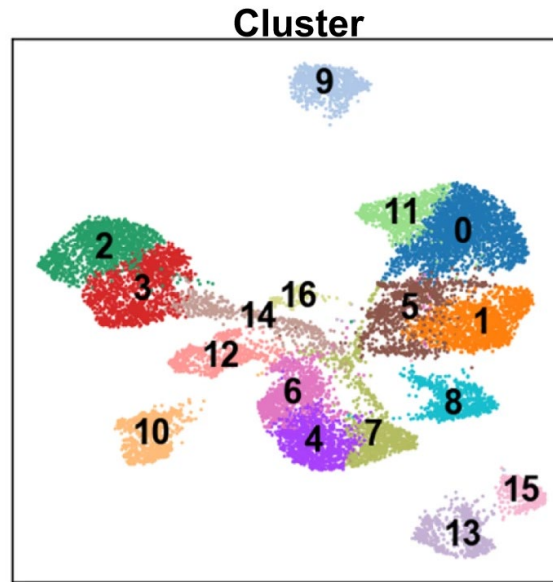
Investigating immune and non-immune cell interactions in head and neck tumors by single-cell RNA sequencing

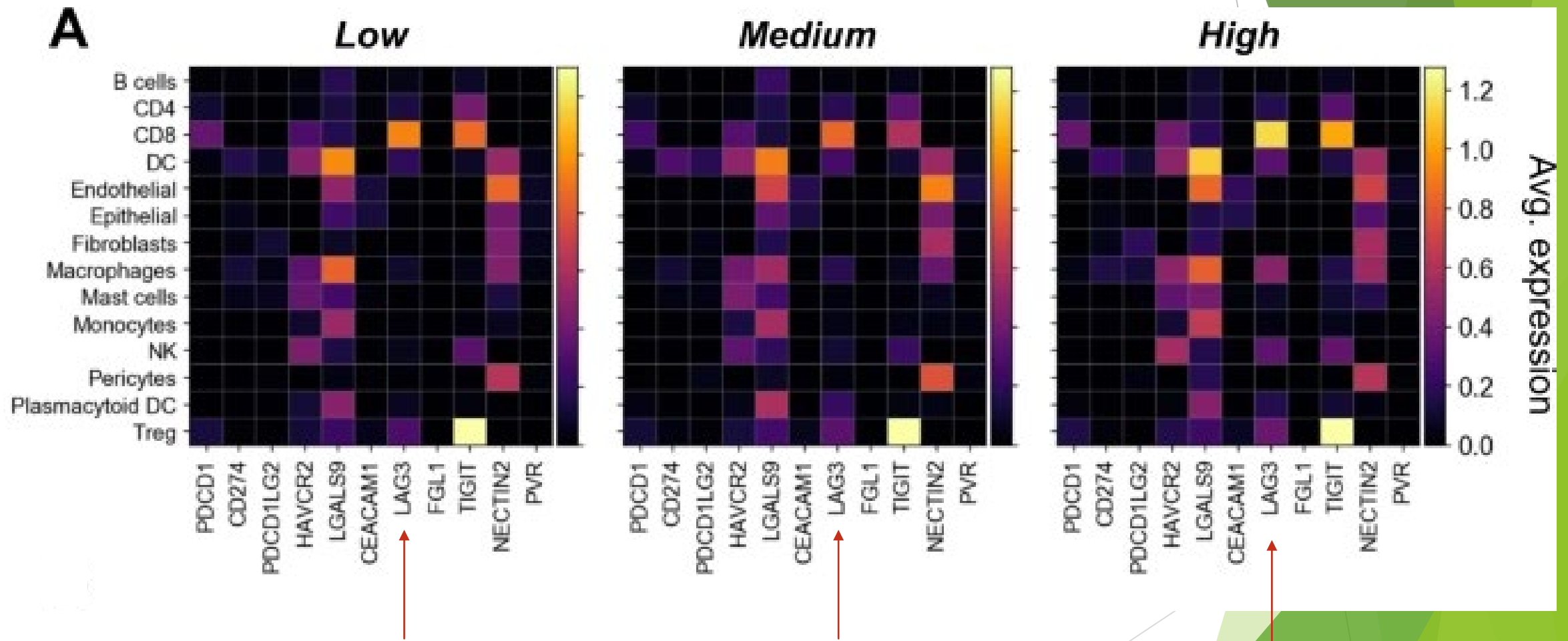
Cornelius H. L. Kürten ^{1,2,3,4,13}, Aditi Kulkarni^{2,3,4,13}, Anthony R. Cillo ^{2,3,5}, Patricia M. Santos^{2,3,4}, Anna K. Roble ^{2,3,4}, Sayali Onkar^{2,3,5,6}, Carly Reeder^{2,3,4}, Stephan Lang¹, Xueer Chen⁷, Umamaheswar Duvvuri⁴, Seungwon Kim⁴, Angen Liu^{4,8}, Tracy Tabib⁹, Robert Lafyatis⁹, Jian Feng^{2,10,11}, Shou-Jiang Gao ^{2,10,11}, Tullia C. Bruno ^{2,3,5}, Dario A. A. Vignali^{2,3,5}, Xinghua Lu ⁷, Riyue Bao^{2,12}, Lazar Vujanovic ^{2,3,4,13} & Robert L. Ferris^{2,3,4,5}✉





UMAP →





Conclusion

- ▶ Viral and cellular heterogeneity remain a challenge in Virology
- ▶ SC technologies allows examination of cell-to-cell variability on the outcome of viral infection
- ▶ Can capture viral diversity and evolution of sequence variation
- ▶ Better understanding of virus infection
- ▶ Allow, drug refinement, personalized medicine

Reference

- ▶ Mertz DR, Ahmed T, Takayama S. Engineering cell heterogeneity into organs-on-a-chip. *Lab Chip*. 2018 Aug 7;18(16):2378-2395. doi: 10.1039/c8lc00413g. PMID: 30040104; PMCID: PMC6081245.
- ▶ Kürten, C.H.L., Kulkarni, A., Cillo, A.R. *et al.* Investigating immune and non-immune cell interactions in head and neck tumors by single-cell RNA sequencing. *Nat Commun* 12, 7338 (2021). <https://doi.org/10.1038/s41467-021-27619-4>
- ▶ Ciuffi A, Rato S, Telenti A. **Single-Cell Genomics for Virology.** *Viruses*. 2016 May 4;8(5):123. doi: 10.3390/v8050123. PMID: 27153082; PMCID: PMC4885078.
- ▶ Rato S, Golumbeanu M, Telenti A, Ciuffi A. Exploring viral infection using single-cell sequencing. *Virus Res*. 2017 Jul 15;239:55-68. doi: 10.1016/j.virusres.2016.10.016. Epub 2016 Nov 2. PMID: 27816430.
- ▶ Svensson, V., Vento-Tormo, R. & Teichmann, S. Exponential scaling of single-cell RNA-seq in the past decade. *Nat Protoc* 13, 599 – 604 (2018). <https://doi.org/10.1038/nprot.2017.149>
- ▶ Zanini F, Robinson ML, Croote D, Sahoo MK, Sanz AM, Ortiz-Lasso E, Albornoz LL, Rosso F, Montoya JG, Goo L, Pinsky BA, Quake SR, Einav S. Virus-inclusive single-cell RNA sequencing reveals the molecular signature of progression to severe dengue. *Proc Natl Acad Sci U S A*. 2018 Dec 26;115(52):E12363-E12369. doi: 10.1073/pnas.1813819115. Epub 2018 Dec 7. PMID: 30530648; PMCID: PMC6310786.

Thank you!

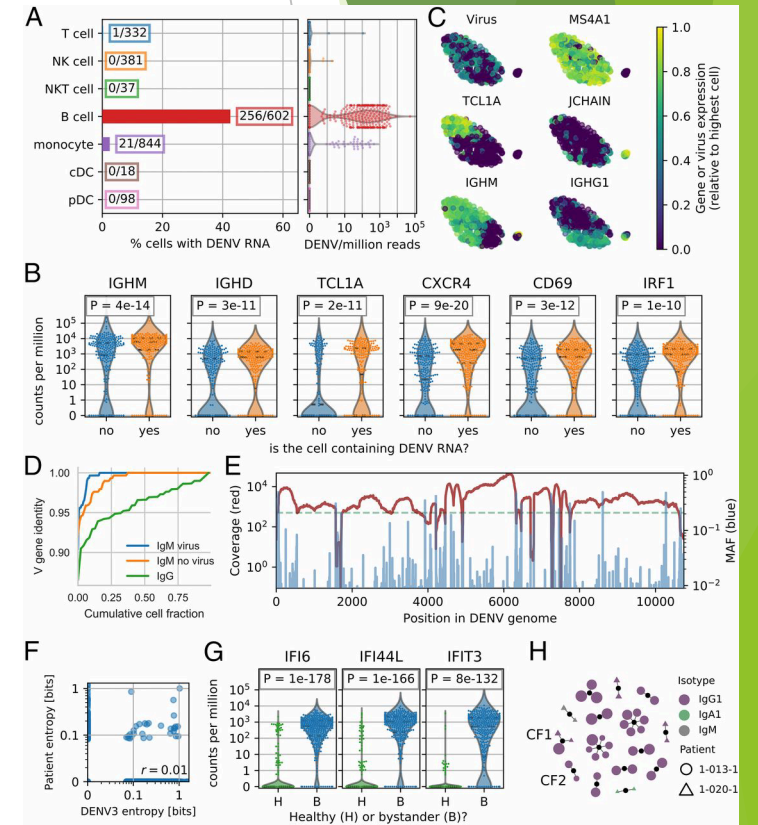
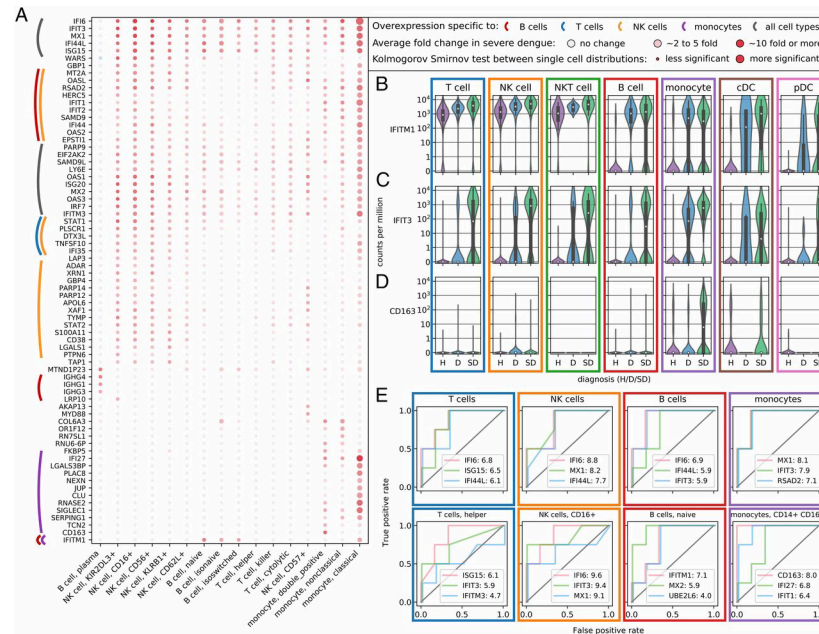
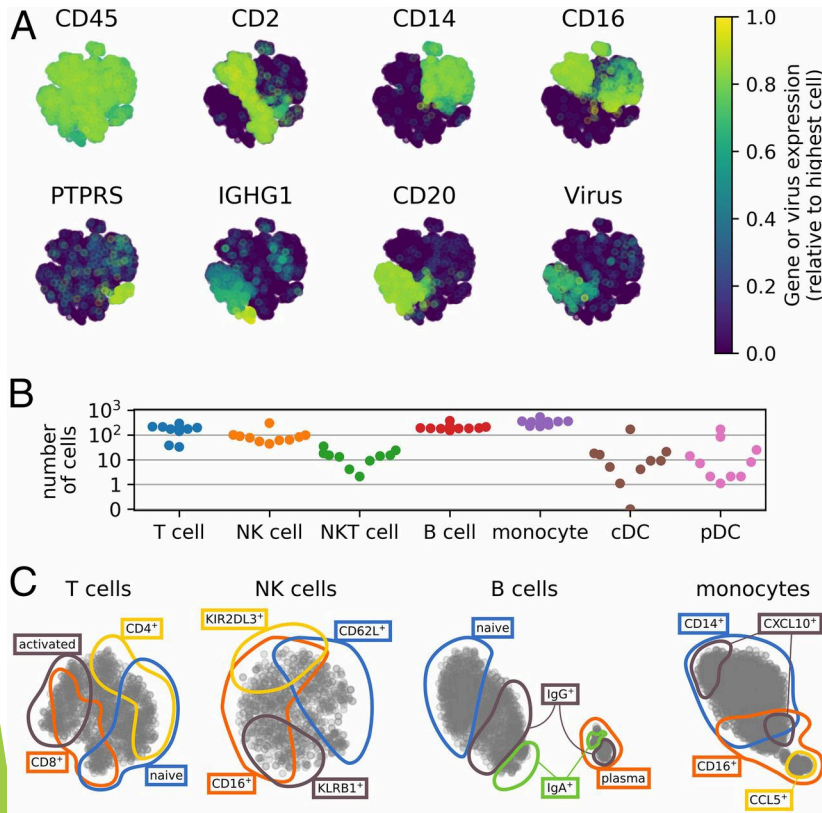
Supplementary (1)

- ▶ DOP-PCR lies in its simplicity and cost-effectiveness, being no more complex than ordinary PCR and requiring little investment in kits or reagents.
- ▶ MALBAC Can sequence large templates, perform single-cell sequencing or sequencing for samples with very limited starting material, Full-amplicon looping inhibits over-representation of templates, reducing PCR bias, but Polymerase is relatively error prone compared to Phi 29, Temperature-sensitive protocol, Genome coverage up to ~90%, but some regions of the genome are consistently underrepresented
- ▶ PMA method allows for full-length transcript coverage to be obtained, but Slight 5' end bias

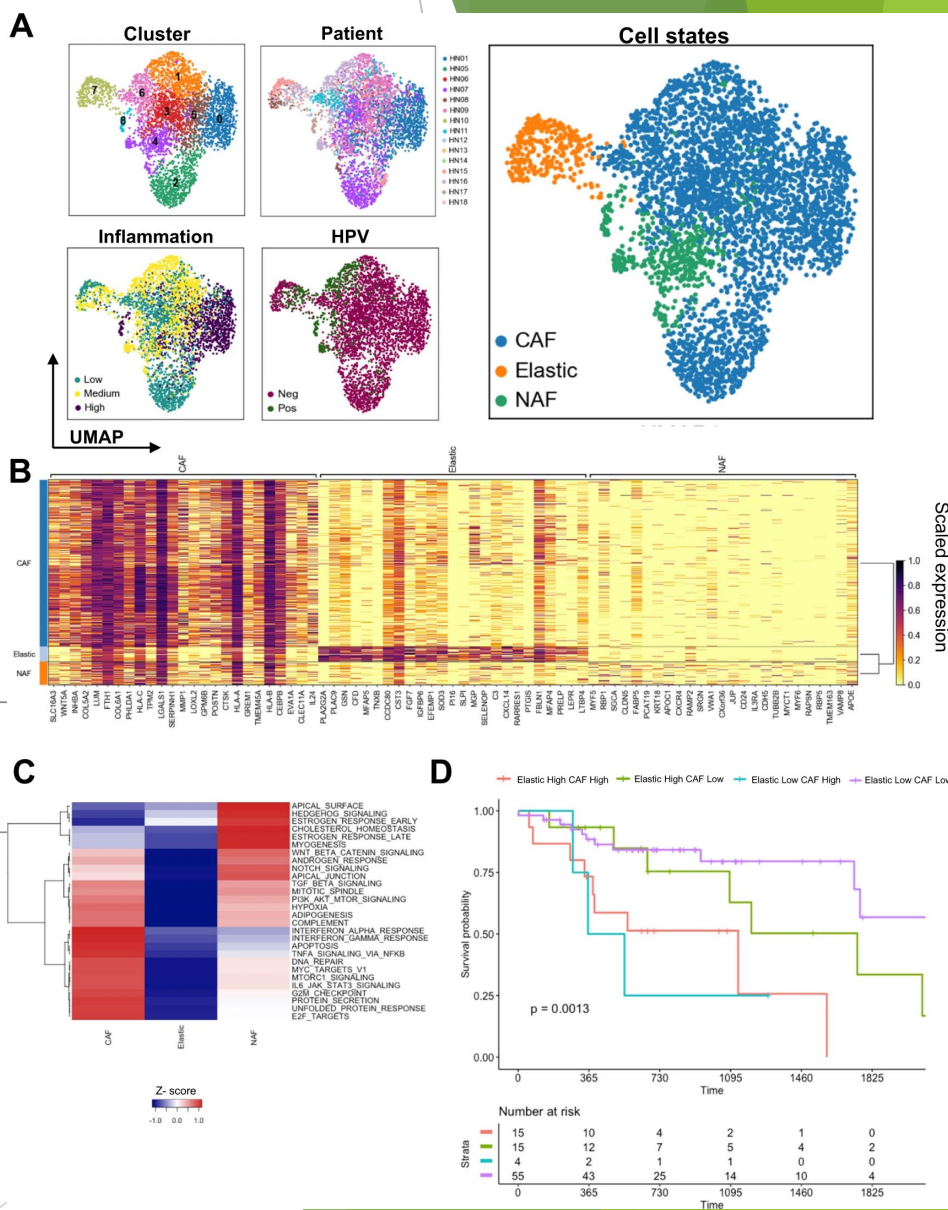
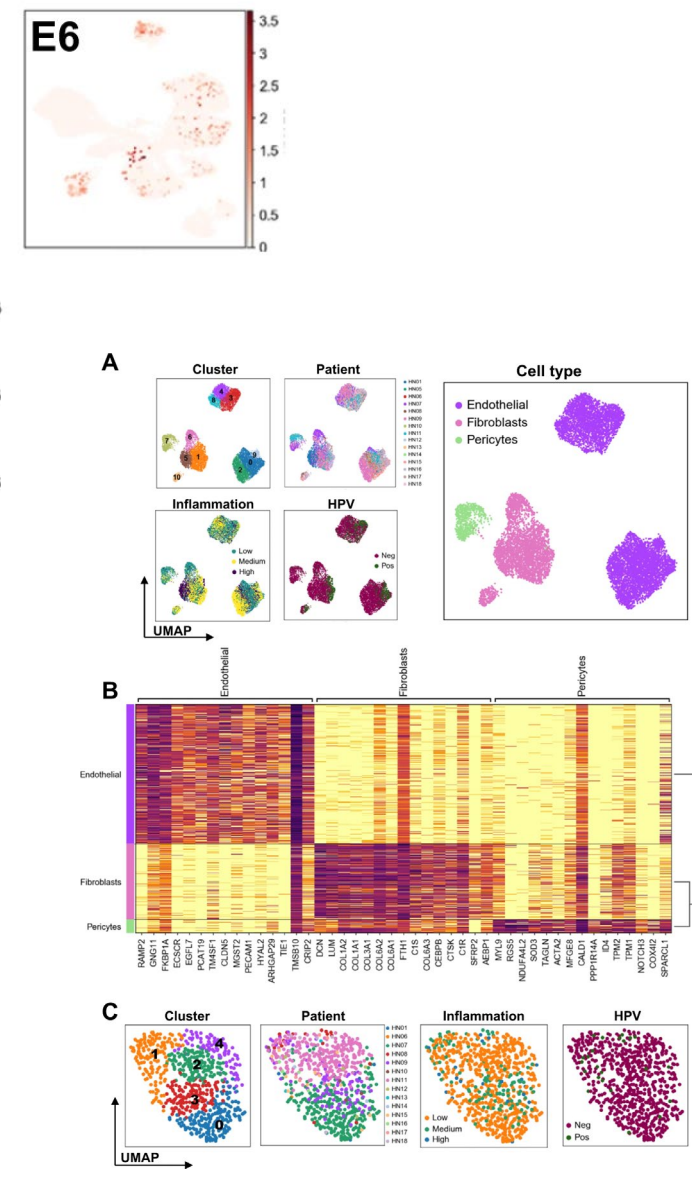
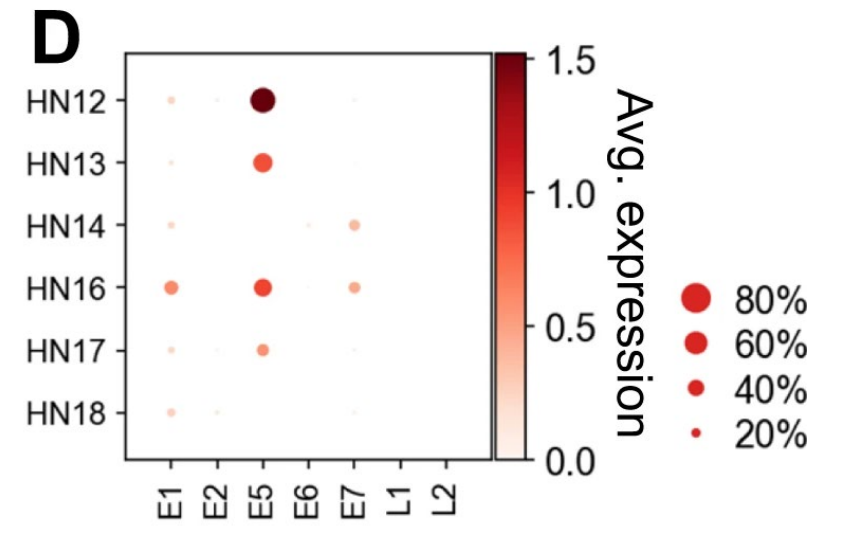
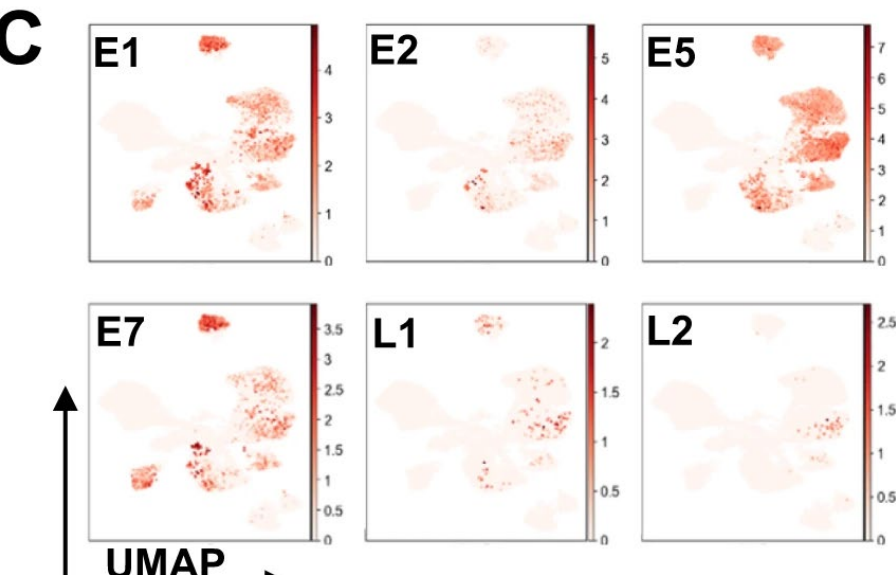
Supplementary (2)

Method	Principle	Coverage	Transcript lengths	Limitaion
CEL-seq	In vitro transcription	49%	Average 1.0 kb	Strong 3' end bias, usually targets the last exons highly
Smart-seq	Modified PCR	Nearly full-length	Average 1.5 kb	Cannot capture partially reverse-transcribed mRNA
PMA	Phi29 DNA polymerase	Full-length	All sizes	Slight 5' end bias

Supplementary (3)



Supplementary (4)



Supplementary (5)

